



Comparative evaluation of air pollution tolerance of plants from polluted and non-polluted regions of Bengaluru

BT Manjunath^{1*}, Jayaram Reddy²

¹Department of Botany, Bharathiar University, Coimbatore, Tamil Nadu, India, ²Department of Botany, St. Joseph's Post Graduate Centre, Bengaluru, Karnataka, India

ARTICLE INFO

Article history:

Received on: October 06, 2018

Accepted on: December 05, 2018

Available online: April 05, 2019

Key words:

Chlorophyll,

Relative Water Content,

Air Pollution Tolerance Index

ABSTRACT

Green vegetation acts as a sink for particulate air pollutants and is considered as ecologically sustainable cost-effective strategies to mitigate ill effects of air pollution in industrialized areas and urban environment. In this context, assessment of air pollution tolerance index (APTI) offers simple tool for screening and identifying plants that tolerate air pollution. Six plants commonly growing in the polluted regions of Bengaluru were selected for assessing the air pollution tolerance response. APTI indices of *Ocimum sanctum*, *Ricinus communis*, *Leucas aspera*, *Lantana camara*, *Bougainvillea spectabilis*, and *Vinca rosea* were assessed and compared with the plants grown in non-polluted sites of Bengaluru to analyze the response to air pollution. APTI correlations with the biochemical and physicochemical parameters were analyzed to identify the important determinants of air pollution tolerance. Among the parameters assessed ascorbic acid content correlated with APTI scores of plants (R, 0.88). Among the plants evaluated, high APTI of >23 was recorded in *B. spectabilis* and *V. rosea*, indicating that these plants are tolerant to air pollutants at the collected sites. *O. sanctum* recorded lowest APTI of 8.77–9.42, indicating that these plants are sensitive to air pollutants and can be used for biomonitoring the air pollution. *L. aspera*, *V. rosea*, and *B. spectabilis* with APTI score of >16 can be used as green belt in polluted areas.

1. INTRODUCTION

Air pollutants comprising particulate matter (PM), vehicular exhaust, and industrial emissions cause adverse health effects in humans, disturb plant ecosystem, and impact globally by altering the atmosphere [1]. Foliar surface of plants acts as a sink for deposition of air pollutants in the urban and industrial environment. Plants that are tolerant to air pollutants and PM are explored as increasingly considered as an eco-sustainable tool for mitigation of air pollution [2]. Screening and identification of plants that are adaptive to the native environment of polluted sites provide ecological restoration strategies to mitigate the impact of air pollution.

Plants differ in their sensitivity to tolerate air pollution and Singh and Rao [3] proposed a formula computing four physicochemical (relative water content [RWC] and leaf extract pH) and biochemical (ascorbic acid and total chlorophyll content) parameters for screening and classifying plants according to their tolerant or sensitive to environmental pollution. Further, air pollution tolerance index (APTI) was applied to classify tree, shrubs, and herbs into sensitive and tolerant based on response of plants to air pollution under laboratory and field experiments [4]. Plants

with APTI index of <11, 12–16, and >17 are considered as sensitive, intermediate tolerance, and tolerant to air pollution [4].

In the present study, physicochemical parameters such as pH of leaf extractives, RWC, and stomatal index were assessed in six plants commonly growing in polluted and the corresponding non-polluted (NP) control sites in Bengaluru city were determined to assess the impact of air pollution. Effect of air pollution on important determinants of plant growth such as chlorophyll and ascorbic acid content was evaluated. APTI was computed and correlated with all the parameters studied to identify important determinant correlating with the tolerance in the polluted and NP areas of Bengaluru.

2. MATERIALS AND METHODS

2.1. Plant Collection and Preparation of Extracts

Fresh leaves of the selected plants from the polluted sites near industries and NP (control sites) were collected. Aqueous extracts were prepared according to standard procedures.

2.2. Biochemical Parameters

2.2.1. pH

About 100 mg of fresh leaves were collected and dried and were homogenized in 10 ml of double distilled water and were filtered and pH of the filtrate containing the leaf extract was examined using calibrated pH meter (pH 4 and pH 9).

*Corresponding Author

Manjunath BT,

Department of Botany, CHRIST (Deemed to be University),
Bengaluru - 560029,

Karnataka, India. Phone: +91-9731903899.

E-mail: manjunath.bt@christuniversity.in

2.2.2. Relative water content

After collection, fresh leaf material was weighed and their initial weight was recorded. Overnight weight of the leaf samples immersed in the distilled water was also recorded after blotting it in dry sheet. Dry weight of the leaves was measured after incubating it in a hot air oven at 70°C and dried weight was reweighed. Relative water content was calculated using standard formula [5].

$$RWC = [(FW - DW) / (TW - DW)] \times 100.$$

Where,

FW indicates fresh weight,

DW indicates dry

TW indicates turgid weight.

2.2.3. Ascorbic acid content

Titrimetric method was used to analyze the ascorbic content of the sample using 2,6-dichlorophenol indophenol dye [6]. 500 mg of collected leaf sample was extracted using 4% oxalic acid and the collected extract was then titrated against the dye. Appearance of pink color was considered as the indicator. Values were subtracted with blank.

2.2.4. Stomatal index

Collected fresh leaves were washed in distilled water and boiled in concentrated nitric acid for 2 min. Epidermis was peeled off from the treated samples and was stained with safranin and the mounted slide was observed in microscope ($\times 10 \times 40$). Upper, middle, and lower region of leaf lamina was examined. Stomatal index was measured using stage and ocular scale. Stomatal index was calculated using the following formula:

$$\text{Stomatal index} = [S / (S + E)] \times 100$$

2.2.5. Total chlorophyll and carotenoids content

Total chlorophyll and carotenoids content was measured using Arnon's method [7]. Freshly collected leaves (500 mg) were ground and extracted with 10 ml of 80% acetone for 15 min and were centrifuged at 2500 rpm for 3 min. The supernatant was collected and the absorbance was measured at 645 nm and 663 nm for chlorophyll a and b and 480 and 510 nm for carotenoids using spectrophotometer. Chlorophyll content was calculated using the formula:

- Total chlorophyll: Chlorophyll a + Chlorophyll b; CTc: $20.2 (D_{645}) + 8.02 (D_{663})$
- Total chlorophyll content: $0.1 \text{ CT} \times (\text{leaf dry weight} / \text{leaf fresh weight})$,
- Carotenoids = $7.6 \times 480 \text{ OD} - 1.49 \times 510 \text{ OD}$.

2.2.6. Calculation of APTI

The air pollution tolerance indices for the selected plants were determined using the following formula [3]:

$$APTI = (A [T + P] + R) / 10.$$

Where,

A indicates ascorbic acid content (mg/gm),

T indicates total chlorophyll (mg/gm),

P indicates pH of the leaf extract,

R indicates relative water content of leaf (%).

2.3. Statistical Analysis

Statistical analysis was carried out using Microsoft Excel Office. Significance of variation in selected plants from polluted and NP areas was measured in independent trials using Student's *t*-test.

3. RESULTS AND DISCUSSION

3.1. Relative Water Content

Relative water content of leaves from polluted and NP source is presented in Figure 1a. Mean RWC of plants from NP areas was 69.42 ± 8.22 (%) which was significantly higher ($P < 0.05$) compared to plants from polluted area (61.28 ± 7.7 %, Figure 1b). Effect on air pollution on water holding capacity of leaves varied with different plants. In *Lantana camara* collected from polluted area, there was maximum 20% reduction in RWC when compared to that of NP area (62.61% in polluted v/s 78.15% in NP). A moderate effect of air pollution assessed by 13–15% reduction in RWC was recorded in *Ocimum sanctum*, *Bougainvillea spectabilis*, and *Leucas aspera*. In *Vinca rosea* and *Ricinus communis* plants from polluted area showed lesser reduction in RWC of 3% (86.29 in polluted and 88.59 in NP) and 7% (53.88 in polluted v/s 58.22 in non-polluted areas), respectively, compared to that from NP area. Plants with higher RWC are having better air pollution, tolerance capacity was reported [8]. Similarly, in our study, *V. rosea* from non-polluted area with highest RWC of 88.59% showed highest APTI of 27.44 [Table 1]. Airborne pollutants are extensively reported to increase the loss of water and nutrients from plant leaves affected by increased protoplasmic permeability, leading to senescence [9]. Higher RWC helps plant in regulating the physiological functions under stress induced by airborne pollutants [10]. Higher water holding capacity of leaves under polluted environment may impart tolerance against the toxic airborne pollutants.

3.2. pH of Leaf Extractives:

The pH of leaf extractives of the leaves from polluted and NP areas is summarized in Figure 2a. The pH in plants from polluted and NP

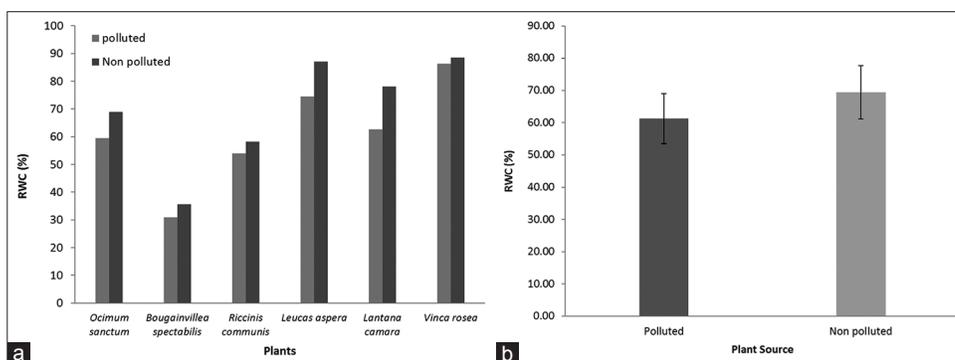


Figure 1: (a and b) Relative water content of selected plants from polluted and non-polluted area

plants ranged from 6.54 to 7.6 and 6.14 to 7.34, respectively. There were significant ($P < 0.05$) variations observed in mean pH of plants from polluted and NP areas (6.84 ± 0.18 in polluted v/s 7.04 ± 0.15 , Figure 2b). However, marked decrease in pH was observed in *L. camara* (-0.40 units) followed by *L. aspera* (-0.31). In *O. sanctum* and *B. spectabilis*, moderate decrease in pH of 0.26 and 0.2 units was recorded. In *R. communis* and *V. rosea* showed similar pH in plants collected from both polluted and NP areas. The presence of acidic pollutants such as SO_2 and NO_2 in air cause lowering of pH. Higher tolerance of plants with higher pH of leaf extractives against pollutants was reported [11]. Higher pH of leaf extractives provides optimal pH for synthesis or reducing activity of important antioxidants such as ascorbic acid, thereby protects enzymes involved in CO_2 fixation cycle and chlorophyll inactivation from the oxidative stress induced by pollutants [12,13]. Photosynthesis efficiency of plants is pH dependent and at lower acidic pH photosynthesis efficiency of plants gets reduced [14]. Thus, the resistance of plants to the air pollutant-induced pH changes could determine the tolerance to air pollution. In our study, in *B. spectabilis* and *L. aspera* with pH of 6.88–6.91 recorded higher APTI [Table 1]; however, no significant [Figure 3] correlation of pH of plants with air pollution tolerance was observed.

3.3. Stomatal Index

The stomatal index of plants from polluted and NP areas is summarized in Figure 4a. The mean stomatal index of plants from polluted environment was found to be significantly ($P < 0.05$) decreased (17.51

± 0.54 in polluted v/s 18.84 ± 0.65 in NP; Figure 4b). Highest effect of pollution was observed in *O. sanctum* (20.50 in NP v/s 17.01 in polluted; 17% reduction in index). In all other plants tested showed slight reduction in stomatal index. Similarly, the effect of air pollution on response to roadside plants with respect to significant reduction in stomatal index in all the plant species growing in polluted area was studied [2]. In contrast, another reported increase in stomatal index in plants from highly polluted sites [15]. Stomatal behavior of plants in response to air pollutants within plant species was reported [16]. Further studies on response of plants to various pollutants are necessary to understand the foliar adaptation of plants exposed to air pollution.

3.4. Total Chlorophyll

Total chlorophyll content of selected plants from polluted and NP areas is summarized in Figure 5a. The mean chlorophyll content of polluted plants was significantly reduced ($P < 0.01$) at $582.30 \pm 21.30 \mu\text{g/g}$ compared to $633.27 \pm 28.60 \mu\text{g/g}$ [Figure 5b]. Highest and lowest total chlorophyll content of $715.12 \mu\text{g/g}$ and $519.48 \mu\text{g/g}$ was recorded in *L. aspera* from NP area and *O. sanctum* from polluted area, respectively. Highest impact of air pollution on total chlorophyll content was recorded in *L. aspera* and *R. communis* where 11–13% reduction in chlorophyll was observed. In *L. aspera*, total chlorophyll content of $620.89 \mu\text{g/g}$ was observed in polluted compared to $715.12 \mu\text{g/g}$ observed in NP in *L. aspera*. Similarly, in *R. communis* plant, total chlorophyll content of $586.69 \mu\text{g/g}$ was observed in polluted compared to $660.26 \mu\text{g/g}$ observed in NP area. In other plants from polluted

Table 1: APTI of selected plants from polluted and non-polluted area

Sample	Stomatal index	Ascorbic acid ($\mu\text{g/g}$)	pH	Total chlorophyll content ($\mu\text{g/g}$)	RWC	APTI
<i>O. sanctum</i> (P)*	17.01	178.65	7.34	519.48	59.45	9.42
<i>Bougainvillea spectabilis</i> (P)	19.25	351.75	6.71	649.35	30.96	23.08
<i>Ricinus communis</i> (P)	16.22	245.72	7.01	586.69	53.88	14.59
<i>Leucas aspera</i> (P)	19.05	301.74	6.57	620.89	74.47	18.94
<i>Lantana camara</i> (P)	17.15	261.62	6.14	594.37	62.61	15.72
<i>Vinca rosea</i> (P)	16.37	487.65	7.25	523.01	86.29	25.87
<i>O. sanctum</i> (NP)*	20.5	157.62	7.6	548.63	68.9	8.77
<i>B. spectabilis</i> (NP)	20.32	226.97	6.91	694.87	35.57	15.93
<i>R. communis</i> (NP)	16.92	205.32	7.02	660.26	58.22	13.71
<i>L. aspera</i> (NP)	19.89	265.98	6.88	715.12	87.06	19.21
<i>L. camara</i> (NP)	18	251.97	6.54	625.5	78.15	15.93
<i>V. rosea</i> (NP)	17.41	328.67	7.31	555.23	88.59	18.50

*P- Polluted, NP-Non-polluted, *Ocimum sanctum*, *B. spectabilis*: *Bougainvillea spectabilis*, *R. communis*: *Ricinus communis*, *L. aspera*: *Leucas aspera*, *L. camara*: *Lantana camara*, *V. rosea*: *Vinca rosea*, RWC: Relative water content, APTI: Air pollution tolerance index

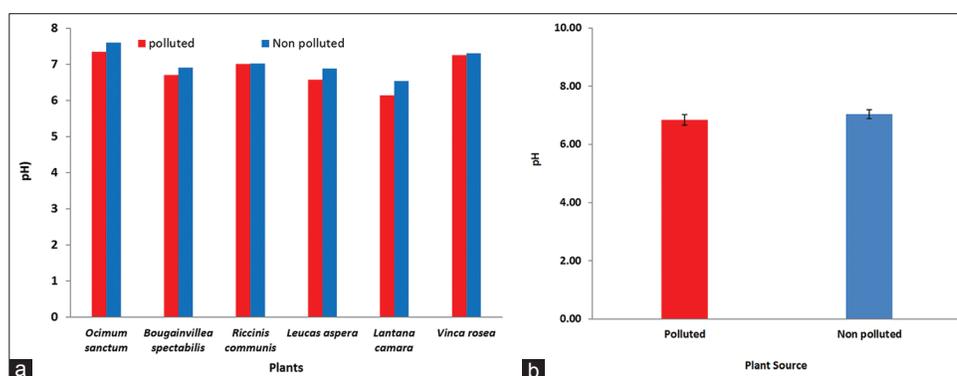


Figure 2: (a and b) pH of selected plants from polluted and non-polluted area

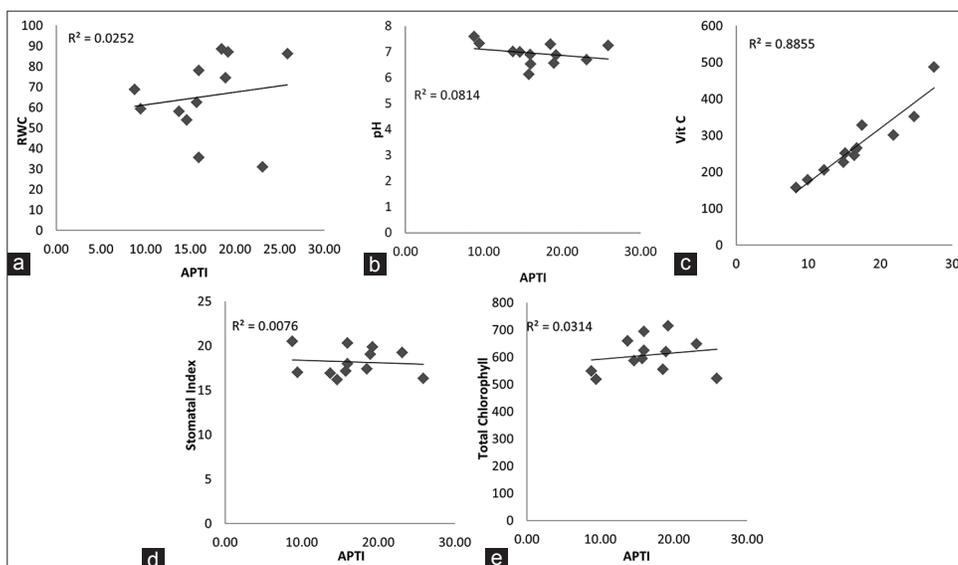


Figure 3: (a-e) Correlation of air pollution tolerance index with physicochemical characteristics of roadside plants

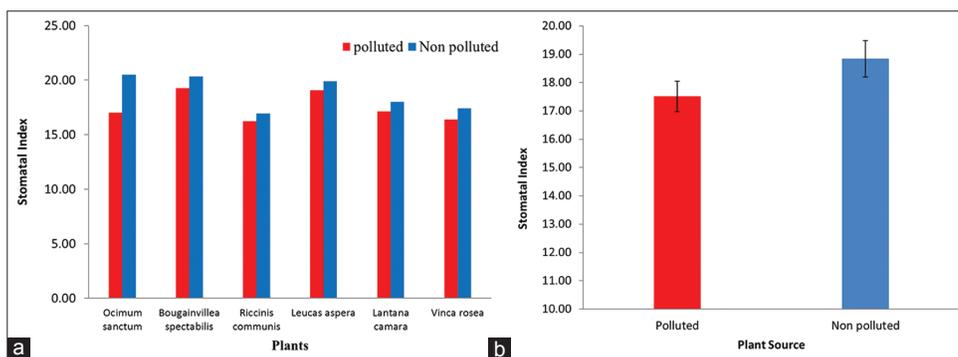


Figure 4: (a and b) Stomatal index of selected plants from polluted and non-polluted area

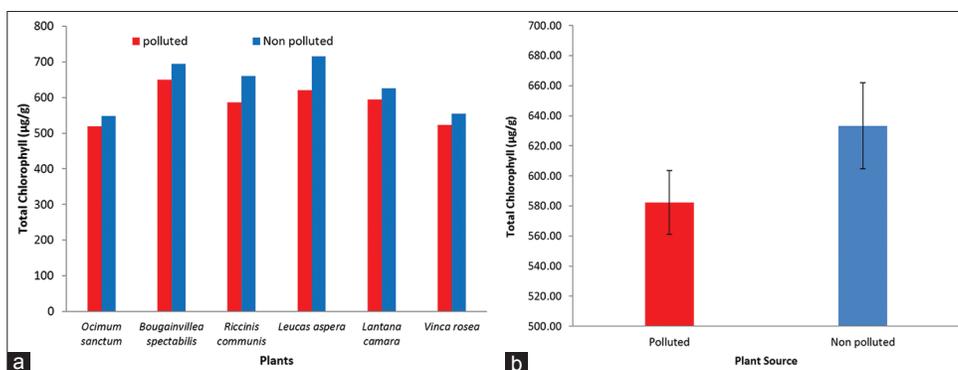


Figure 5: (a and b) Total chlorophyll content of selected plants from polluted and non-polluted area

area, 5–7% reduction in total chlorophyll content was noted. Total chlorophyll content is an important biochemical factor that impacts photosynthetic activity which determines the plant growth. Total chlorophyll content is considered frequently to evaluate the impact of air pollution on plants [4]. Several studies have reported reduction in total chlorophyll content in plants exposed to air pollutants. Effect of PM in polluted area on reduction of chlorophyll content at the enzyme level and correlated with increased activity of chlorophyllase enzyme activity was studied [17]. Reduction in total chlorophyll content to reduce gaseous exchange due to blockage of stomatal opening in

response to air pollutants was analyzed [18]. The study on inverse relationship of total chlorophyll content with dust accumulation on leaves in polluted environment was reported [19]. In our study, the reduced total chlorophyll content could be due to the reduced stomatal index. Comparative analysis of the correlation of total chlorophyll content with air pollution tolerance indices showed no significant correlation [Figure 3c]. Further, detailed correlation analysis on influence of various physicochemical factors on photosynthetic pigments synthesis and activity should help in identifying adaptive plants with sustained growth in polluted environments.

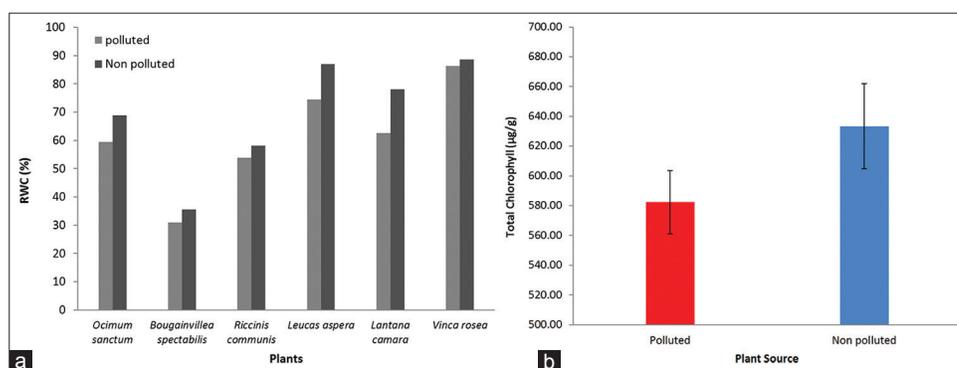


Figure 6: (a and b) Ascorbic acid content of selected plants from polluted and non-polluted area

3.5. Ascorbic Acid Content

Ascorbic acid content of selected plants from polluted and NP areas is summarized in Figure 6a. The mean ascorbic acid content of polluted plants was significantly increased ($P < 0.05$) at $304.52 \pm 7.04 \mu\text{g/g}$ compared to $239.42 \pm 23.69 \mu\text{g/g}$ in NP areas [Figure 6b]. Highest and lowest total ascorbic acid content of $487.65 \mu\text{g/g}$ and $157.62 \mu\text{g/g}$ recorded in *V. rosea* from polluted area and *O. sanctum* from NP area, respectively. Highest impact of air pollution on total ascorbic acid content was recorded in *B. spectabilis* ($351.75 \mu\text{g/g}$ in polluted v/s 226.97 in NP area) and *V. rosea* (487.65 in polluted vs. $328.67 \mu\text{g/g}$ in NP area) with 55% and 48% increase, respectively, in ascorbic acid content was observed. Moderate impact of air pollution on ascorbic acid content was observed in *R. communis* (245.72 in polluted v/s $205.32 \mu\text{g/g}$ in NP area), *O. sanctum* (157.62 in polluted v/s $178.65 \mu\text{g/g}$ in NP area), and *L. aspera* (301.74 in polluted v/s $265.98 \mu\text{g/g}$ in NP area) where 13–20% increase in ascorbic acid content was recorded [Table 1]. In *L. camara*, 4% increase in ascorbic acid content was observed (261.62 in polluted v/s $251.97 \mu\text{g/g}$ in NP area). Several researchers have studied and reported positive correlation of air pollution on ascorbic acid content [19-21].

Ascorbic acid is an important antioxidant which provides resistance from stress in plants by neutralizing the free radicals generated that can affect the biochemical and physiological activity [22,23]. Thus, ascorbic acid is an important determinant that decides the plants ability to nullify and withstand the toxic effects of air pollutants [24,4]. Correlation of air pollution tolerance indices of plants with all the parameters revealed positive correlation with only ascorbic acid content ($R^2=0.88$, Figure 3e). A study reported similar findings of positive correlation of APTI with ascorbic acid content of plants [19].

3.6. APTI

APTI of selected plants from polluted and NP areas is summarized in Table 1. APTI of plants from polluted area ranged from 9.42 to 25.87. APTI of plants from non-polluted area ranged from 8.77 to 19.21. Lowest APTI was recorded for *O. sanctum* selected from both polluted and non-polluted area at 9.42 and 8.77, respectively, indicating that it is sensitive to air pollution. Highest APTI of 25.87 was recorded for *V. rosea* followed by *B. spectabilis* with APTI of 23.08. Moderate APTI of between 12 and 16 was observed in different plants selected from polluted sites in the order of *L. aspera* (APTI 18.94) > *L. camara* (APTI 15.72) > *R. communis* (APTI 14.59). Among the plants selected from non-polluted areas, the APTI score was in the order of *L. aspera* (APTI 19.21) > *V. rosea* (APTI 18.50) > *B. spectabilis* and *L. camara* (APTI 15.93) > *R. communis* (APTI 13.71) > *O. sanctum* (APTI 8.77). APTI indices in plants reveal that the plants vary in their response

to environmental pollution and their tolerance to air pollution is determined by their ability to undergo physicochemical adaptation to either prevent pollutants or mitigate the stress induced by pollutants through the antioxidant. Ascorbic acid content in the plants can be used for screening of plants with favorable tolerance to air pollution. The findings indicate that the plants *B. spectabilis* and *V. rosea* can be selected for growing in polluted environments of Bengaluru.

4. CONCLUSIONS

In the present study, comparative analysis of APTI indices of six common plants growing in the polluted areas and NP control area in Bengaluru was analyzed as a biomonitoring tool to assess the response of plants to air pollution-induced stress. The study findings reveal that *V. rosea* and *B. spectabilis* can be effectively used for the amelioration of air pollution effects. Further, efforts to evaluate these plants to ameliorate air pollutants at heavily polluted urban environments should facilitate exploring these plants for restoration of green urban ecosystem.

5. CONFLICTS OF INTEREST

All the authors declare that there are no conflicts of interest with respect to the data provided and the authors do not claim for any funding sources.

6. REFERENCES

1. Raabe OG. Respiratory exposure to air pollutants. In: Swift DL, Foster WM, editors. Air Pollutants and the Respiratory Tract. New York, USA: Marcel Dekker Inc.; 1999.
2. Prabhat KR. Biodiversity of roadside plants and their response to air pollution in an Indo-Burma hotspot region: Implications for urban ecosystem restoration. J Asia Pac Biodivers 2016;9:47-55.
3. Singh SK, Rao DN. Evaluation of Plants for Their Tolerance to Air Pollution. In: Proceedings Symposium on Air Pollution Control, Indian Association for Air Pollution Control; 1983. p. 218-224.
4. Singh SK, Rao DN, Agrawal M, Pandey J, Narayan D. Air pollution tolerance index of plants. J Environ Manag 1991;32:45-55.
5. Singh A. Practical Plant Physiology. New Delhi: Kalyari Publishers; 1977.
6. Sadasivam S, Balasubraminan T. In: Practical Manual in Biochemistry. Coimbatore: Tamil Nadu Agricultural University; 1987.
7. Aron DI. Copper enzymes in isolated chloroplasts. Polyphenol oxidase in *Beta vulgaris*. Plant Physiol 1949;24:1-15.
8. Paulsamy S, Sivakumar R, Latha N. Evaluation of air pollution tree species in Coimbatore city. J Ecol Res Biocon 2000;1:20-3.
9. Keller T, Schwager H. Air pollution and ascorbic acid. Eur J For

- Pathol 1977;7:338-50.
10. Tsega YC, Prasad AD. Variation in air pollution tolerance index and anticipated performance index of roadside plants in Mysore, India. *J Environ Biol* 2014;35:185-90.
 11. Agrawal AL. Air Pollution Control Studies and Impact Assessment of Stack and Fugitive Emissions from CCI Akaltara Cement Factory. Project Report, Project Sponsored by M/s. CCI Akaltara Cement Factory. Nagpur: NEERI; 1988.
 12. Tanaka K, Otsubo T, Kondo N. Participation of hydrogen peroxide in the inactivation of Calvin-cycle SH enzymes in SO₂-fumigated spinach leaves. *Plant Cell Physiol* 1982;28:1009-18.
 13. Choudhary CS, Rao DN. Study of some factors in plants controlling their susceptibility to SO₂ pollution. *Proc Indian Natl Sci Acad* 1977; 43:236-41.
 14. Yan-ju L, Hui D. Variation in air pollution tolerance index of plant near a steel factory: Implications for landscape-plant species selection for industrial areas. *WSEAS Trans Environ Dev* 2008;1:24-30.
 15. Gostin I. Air pollution effects on the leaf structure of some *Fabaceae* species. *Notulae Bot Horti Agrobotanici Cluj-Napoca* 2009;37:57-63.
 16. Robinson MF, James H, Mansfield TA. Disturbances in stomatal behaviour caused by air pollutants. *J Exp Bot* 1998;49:461-9.
 17. Prajapati SK. Ecological effect of airborne particulate matter on plants. *Environ Skeptism Crit* 2012;1:12-22.
 18. Leghari SK, Zaidi MA. Effect of air pollution on the leaf morphology of common plant species of Quetta city, Pakistan. *Pak J Bot* 2013;45:447-54.
 19. Bharti SK, Kumar D, Anand S, Poonam, Barman SC, Kumar N. Characterization and morphological analysis of individual aerosol of PM10 in urban area of Lucknow, India. *Micron* 2017;103:90-8.
 20. Govindaraju M, Kumar RS, Muthukumaran VR, Visvanathan P. Identification and evaluation of air-pollution tolerant plants around lignite-based thermal power station for greenbelt development. *Environ Sci Pollut Res* 2012;19:1210-23.
 21. Pandey AK, Pandey M, Mohan S, Tiwary M, Tripathi BD. Air pollution tolerance index and anticipated performance index of some plant species for development of urban forest. *Urban for Urban Green* 2015;14:866-71.
 22. Keller T. The electrical conductivity of Norway spruce needle diffusate as affected by air pollutants. *Tree Physiol* 1986;1:85-94.
 23. Conklin PL. Recent advances in the role and biosynthesis of ascorbic acid in plants. *Plant Cell Environ* 2001;24:383-94.
 24. Aghajanzadeh T, Hawkesford MJ, De Kok LJ. Atmospheric H₂S and SO₂ as sulfur sources for *Brassica juncea* and *Brassica rapa*: Regulation of sulfur uptake and assimilation. *Environ Exp Bot* 2016; 124:1-10.

How to cite this article:

Manjunath BT, Reddy J. Comparative evaluation of air pollution tolerance of plants from polluted and non-polluted regions of Bengaluru. *J App Biol Biotech.* 2019;7(03):63-68. DOI: 10.7324/JABB.2019.70312